

# Combination Therapy of Intra-bony Periodontal Defects Using an Er:YAG Laser and Enamel Matrix Protein Derivative: A Case Series

Frank Schwarz<sup>a</sup>, Anton Sculean<sup>b</sup>, Katrin Bieling<sup>c</sup>, Jürgen Becker<sup>d</sup>, George E. Romanos<sup>e</sup>

<sup>a</sup> Assistant Professor, Department of Oral Surgery, Heinrich Heine University, Düsseldorf, Germany.

<sup>b</sup> Professor and Chairman, Department of Periodontology, Radboud University Medical Center, Nijmegen, The Netherlands.

<sup>c</sup> Research Associate, Department of Oral Surgery, Heinrich Heine University, Düsseldorf, Germany.

<sup>d</sup> Professor and Chairman, Department of Oral Surgery, Heinrich Heine University, Düsseldorf, Germany.

<sup>e</sup> Assistant Professor, Department of Periodontology and Implant Dentistry, New York University College of Dentistry, New York, NY, USA.

**Purpose:** The aim of the present series of case reports was to evaluate the combination therapy of intra-bony periodontal defects using an Er:YAG laser (ERL) and enamel matrix protein derivative (EMD).

**Materials and Methods:** Fifteen patients with chronic periodontitis, each of whom displayed one intra-bony defect, were randomly treated with access flap surgery and root surface/defect debridement using ERL (16.5 J/cm<sup>2</sup>) followed by the application of EMD. The following clinical parameters were recorded at baseline and at 6 months: plaque index (PI), bleeding on probing (BOP), probing pocket depth (PD), gingival recession (GR), and clinical attachment level (CAL).

**Results:** Healing was uneventful in all patients. At 6 months after therapy, the sites treated with ERL and EMD showed a mean PD reduction from 8.5 ± 1.1 mm to 4.7 ± 0.8 mm and a mean CAL change from 10.5 ± 1.4 mm to 7.6 ± 1.4 mm (p < 0.001).

**Conclusion:** Within the limits of this case series, it may be concluded that the combination of ERL and EMD may improve clinical healing of intra-bony periodontal defects over a period of 6 months.

**Keywords:** case series, enamel proteins, intra-bony defects, lasers, periodontal regeneration, therapeutic use.

*J Oral Laser Applications 2006; 6: 123-131.*

According to the cause-related concept of periodontal therapy, the main objective of treatment is to control infection and thereby arrest disease progression.<sup>1</sup> Ideally, periodontal therapy does not only include arresting the disease but also regeneration of the periodontal attachment, including cementum, a functionally oriented periodontal ligament, and alveolar

bone.<sup>2</sup> Several treatment modalities, such as the use of different types of bone grafts, root surface demineralization, guided tissue regeneration (GTR), or the application of growth factors have been employed with varying degrees of success in order to predictably accomplish this goal.<sup>3-7</sup> Enamel matrix derivative (EMD) has also been introduced as an alternative modality in



regenerative periodontal treatment.<sup>8</sup> The rationale for the clinical use of EMD is the observation that enamel matrix proteins (EMPs) are deposited along the surface of developing tooth roots prior to cementum formation.<sup>8</sup> It has been suggested that EMPs are involved in the formation of acellular extrinsic fiber cementum and may trigger differentiation of progenitor cells into cementoblasts.<sup>8</sup> Recent data have also indicated that EMD may contain additional mitogenic factors, such as TGF, and BMP-like growth factors, that stimulate fibroblastic proliferation and contribute to the induction of biomineralization during periodontal regeneration.<sup>9-12</sup> Indeed, histological findings from both animals<sup>13</sup> and humans<sup>14-16</sup> have shown that the application of EMD onto a debrided root surface may promote periodontal regeneration. Furthermore, clinical results have shown that treatment of intrabony defects with EMD may result in clinical outcomes comparable to those following GTR therapy.<sup>17,18</sup> Moreover, clinical trials have reported improved gains in clinical attachment levels compared to access flap surgery alone.<sup>18-20</sup>

Usually, periodontally diseased root surfaces are debrided using hand instruments. However, the formation of a smear layer after mechanical scaling and root planing (SRP) has been reported to be detrimental to periodontal tissue healing, as it may inhibit reattachment of cells to the root surface.<sup>21,22</sup> In order to improve the biocompatibility, root surface conditioning with various substances, such as ethylene diamine tetraacetic acid gel (EDTA) at neutral pH, citric and ortho-phosphoric acids, has been proposed.<sup>21-24</sup> The biological rationale of using EDTA for root surface conditioning is based on findings from experimental studies, indicating that EDTA operating at neutral pH appeared to effectively remove the smear layer produced by SRP and to selectively remove mineral from the dentin or cementum surface, exposing a collagenous matrix.<sup>21,22,25,26</sup> In contrast, etching with citric and phosphoric acids appeared to remove not only the mineral component but also the collagenous matrix.<sup>21,22,25,26</sup> The exposure of a collagenous matrix may also be favorable for retention of biologically active substances, such as EMD.<sup>27</sup>

In recent years, the use of laser radiation has also been expected to serve as an alternative or adjunctive treatment to conventional periodontal therapy. Various hypothetically advantageous characteristics, such as hemostatic effects, improved calculus removal, and bactericidal effects against periodontopathic pathogens, might lead to improved treatment outcomes.<sup>28</sup> Close attention has been paid to the clinical applicability of the Er:YAG laser (ERL) with a wavelength of 2.940 nm

in the near infrared spectrum.<sup>29</sup> This laser system effectively removes calculus from periodontally diseased root surfaces without producing thermal side effects in adjacent tissues.<sup>30-32</sup> The absence of thermal damage was most likely due to the optical characteristics of its wavelength of 2.940 nm, since the ERL laser theoretically has a 10 and 15,000 to 20,000 times higher absorption coefficient of water than the CO<sub>2</sub> and the Nd:YAG lasers, respectively.<sup>33,34</sup> Additionally, recently published studies have reported a lack of cementum removal when laser instrumentation was performed under in vivo conditions.<sup>31,32,35</sup> Furthermore, several studies have reported antimicrobial effects against periodontopathic bacteria and the removal of lipopolysaccharides by ERL radiation.<sup>36-40</sup> These findings suggest that root surface debridement and detoxification using ERL may also facilitate a precipitation of EMD. Indeed, preliminary results from a pilot study have shown that the combination therapy ERL+EMD resulted in statistically significant clinical attachment level gains.<sup>41</sup> However, data on clinical outcomes following treatment of intrabony defects with ERL+EMD are still limited.

Therefore, the aim of the present series of case reports was to evaluate the combination therapy of intrabony periodontal defects using ERL and EMD.

## MATERIALS AND METHODS

### Study Population

Fifteen patients suffering from chronic periodontitis were included in the present series of case reports. The patient population comprised 7 men and 8 women (mean age: 49 ± 12 years). The inclusion criteria were: (1) no systemic diseases that could influence the outcome of the therapy; (2) a good level of oral hygiene (plaque index <1<sup>42</sup>); (3) compliance with the maintenance program; and (4) presence of one intrabony defect with a probing depth ≥ 6 mm and an intrabony component of ≥ 3 mm as detected on radiographs.

Patients who reported smoking only occasionally were not considered as smokers.<sup>43</sup> According to the given definition, there were 6 smokers included in the present study.

Three months prior to surgery, each patient was given thorough oral hygiene instructions, and full mouth supra- and subgingival scaling using ERL (KEY2/KEY3, KaVo, Biberach, Germany).

## Clinical Measurements

The following clinical parameters were assessed 1 week prior to and 6 months after the surgical procedure using a periodontal probe (PCP 12, Hu-Friedy; Leimen, Germany): plaque index (PI),<sup>42</sup> bleeding on probing (BOP), probing depth (PD), gingival recession (GR), and clinical attachment level (CAL). The measurements were made at 6 sites per tooth: mesiovestibular (mv), midvestibular (v), distovestibular (dv), mesiolingual (ml), midlingual (l), and distolingual (dl) by one blinded and calibrated investigator. Five patients, each showing two pairs of contralateral teeth (single- and multi-rooted) with probing depths  $\geq 6$  mm at at least one site of each tooth, were used to calibrate the examiner. The examiner evaluated the patients on 2 separate occasions, 48 h apart. Calibration was accepted if measurements at baseline and at 48 h were within a millimeter 90% or more of the time. The cemento-enamel junction (CEJ) was used as the reference point. In cases where the CEJ was not visible, a restoration margin was used for these measurements. The study reports only measurements at the same deepest point of the selected defect. Prior to surgery and 6 months postoperatively, periapical radiographs were taken using the long-cone paralleling technique.

## Surgical Procedure

All surgical procedures were performed under local anesthesia. Following intracrevicular incisions, full-thickness mucoperiosteal flaps were raised vestibularly and orally. Both the removal of the granulation tissue from the intrabony defects and debridement of root surfaces was performed using the ERL (KEY2/KEY3 without feedback system; KaVo) device emitting a pulsed infrared radiation at a wavelength of 2.94  $\mu\text{m}$ . The laser beam was guided under water irrigation with a specially designed periodontal handpiece and a chisel-shaped glass fiber tip (size 0.5 x 1.65 mm) (2061, KaVo). Laser parameters were set at 160 mJ/pulse and 10 pulses/s, and pulse energy at the tip was approximately 136 mJ/pulse (16.5 J/cm<sup>2</sup>).<sup>44-46</sup> Debridement of the root surfaces was performed from coronal to apical in parallel paths with an inclination of the fiber tip of 15 to 20 degrees<sup>47</sup> to the root surface. EMD (Emdogain, Straumann; Waldenburg, Switzerland) was applied to the root surfaces and into the defects according to the instructions given by the manufacturer. Finally, the flaps were repositioned coronally and closed with vertical or horizontal mattress sutures (Figs

1a to 1h). During surgery, the following measurements were made: distance from the CEJ to the bottom of the defect (CEJ-BD) and distance from the CEJ to the most coronal extension of the alveolar bone crest (CEJ-BC). The intrabony component (INTRA) of the defects was defined as (CEJ-BD) minus (CEJ-BC).

## Postoperative Care

Postoperative care consisted of 0.2% chlorhexidine rinses twice a day for 4 weeks. The sutures were removed 10 days after surgery. Recall appointments were scheduled every second week during the first 2 postoperative months, and once a month for the rest of the observation period. Neither probing nor subgingival instrumentation was performed during the first six months after surgery.

## Statistical Analysis

A software package (SPSS 13.0, SPSS; Chicago, IL, USA) was used for the statistical analysis. The primary outcome variable was CAL. In the calculations, only the deepest site per tooth was included. The data rows were examined with the Kolmogorow-Smirnow test for normal distribution. Accordingly, the paired t-test was used to statistically evaluate the changes from baseline to 6 months.

## RESULTS

The depth and configurations of the intrabony components as assessed during surgery is presented in Table 1. Before surgery, one tooth received additional root canal treatment due to a combined periodontal and endodontic lesion.

Postoperative healing was considered as generally uneventful. Minor complications were related to the usual postoperative swelling and occurred within the first days after surgery. Neither allergic reactions nor suppuration or abscesses were observed in any of the patients.

Mean clinical parameters at baseline and after 6 months are summarized in Table 2. Mean PI remained low throughout the study period. In general, ERL+EMD resulted in statistically significant improvements of all clinical parameters investigated. In particular, BOP improved statistically significantly from 100% at baseline to 33% after 6 months ( $p < 0.001$ ). Mean PD changed



**Fig 1a** Clinical situation at baseline.



**Fig 1b** Acute periodontal inflammation on the lingual aspect of tooth 32.



**Fig 1c** Bilateral intrasulcular incisions and mobilization of mucoperiosteal flaps.



**Fig 1d** Two-wall intrabony defect on the lingual aspect of tooth 32.



**Fig 1e** Removal of granulation tissue and root surface debridement using ERL.



**Fig 1f** Removal of a thin blood film from the root surface.



Fig 1g Application of EMD.



Fig 1h Wound closure.

Table 1 Baseline defect characteristics in mm (mean ± SD) (n = 15 patients)						
Treatment	CEJ-BD	CEJ-BC	INTRA	1-2-wall	2-wall	3-wall
ERL+EMD	12.5 ± 1.4	8.7 ± 1.6	3.8 ± 0.9	4	9	2

Table 2 Clinical parameters (mean ± SD) at baseline and after 6 months (n = 15 patients)				
	Baseline	6 months	Differencep	P value
PI	0.9 ± 0.7	0.6 ± 0.6	0.3 ± 0.2	> 0.05
BOP (%)	100%	33%	67%	< 0.001
PD (mm)	8.5 ± 1.1 mm	4.7 ± 0.8 mm	3.9 ± 0.9 mm	< 0.001
GR (mm)	2.0 ± 0.9 mm	2.9 ± 1.0 mm	0.9 ± 0.6 mm	< 0.001
CAL (mm)	10.5 ± 1.4 mm	7.6 ± 1.4 mm	3.0 ± 1.2 mm	< 0.001

from  $8.5 \pm 1.1$  mm at baseline to  $4.7 \pm 0.8$  mm after 6 months ( $p < 0.001$ ), and mean CAL changed from  $10.5 \pm 1.4$  mm at baseline to  $7.6 \pm 1.4$  mm after 6 months ( $p < 0.001$ ) (Figs 1i and 1j). Radiological observation revealed a decreased translucency within the intrabony component of each defect investigated (Figs 1k, 1l, and 2).

The sample size was too small to draw any conclusions regarding the effects of smoking on clinical outcomes.

## DISCUSSION

The findings of the present case series have shown that treatment of intrabony periodontal defects with ERL+EMD may lead to clinically important and statistically significant PD reductions and CAL gains. The fact that all defects treated in this study healed uneventfully suggests that this treatment modality was well tolerated. The present results corroborate recent findings from a pilot study evaluating the clinical use of ERL+EMD and SRP+EDTA+EMD for the treatment of intrabony periodontal defects.<sup>41</sup> In particular, the sites treated with ERL+EMD showed a reduction in mean



**Fig 1i** Clinical situation after 6 months.



**Fig 1j** Healthy periodontal conditions also at the lingual aspect.



**Fig 1k** Preoperative radiograph.



**Fig 1l** Postoperative radiograph at 6 months.

PD from  $8.6 \pm 1.2$  mm at baseline to  $4.6 \pm 0.8$  mm after 6 months, and a change in mean CAL from  $10.7 \pm 1.3$  mm at baseline to  $7.5 \pm 1.4$  mm after 6 months. In the group treated with SRP+EDTA+EMD, the mean PD was reduced from  $8.1 \pm 0.8$  mm at baseline to  $4.0 \pm 0.5$  mm after 6 months, and the mean CAL changed from  $10.4 \pm 1.1$  mm at baseline to  $7.1 \pm 1.2$  mm after 6 months. In both groups, the differences between baseline and 6 months were statistically significant. However, the differences between the groups were not statistically significant.

The finding that treatment of intrabony periodontal defects with EMD may result in the short term (up to 1 year) in statistically significant improvements in PD and CAL compared to baseline is in agreement with already reported results.<sup>17,19,20,48</sup> In particular, Hejil et al<sup>19</sup> reported a mean CAL gain of 2.1 mm after 8 months (baseline CAL: 9.4 mm; INTRA: 4.8 mm). There was a statistically significant difference between EMD- and placebo-treated sites. Similarly, Pontoriero et al<sup>20</sup> reported a mean CAL gain of 2.9 mm for EMD-treated sites after 1 year with a statistically significant



**Fig 2a** Preoperative radiograph.



**Fig 2b** Postoperative radiograph at 6 months. Before surgery, tooth 33 received an additional root canal treatment due to a combined periodontal and endodontic lesion.

difference between EMD- and placebo-treated sites (baseline CAL: 9.1 mm; INTRA 4.2 mm). Froum et al<sup>48</sup> reported a 4.26 mm CAL gain (baseline CAL: not reported; INTRA: 5.63 mm), and Sculean et al<sup>17</sup> reported a 3.4 mm CAL gain (baseline CAL: 10.6; INTRA 3.8 mm). In these clinical trials, the most frequently employed root conditioning agent was 24% EDTA.<sup>17,20,48</sup>

Furthermore, the present results obtained with ERL+EMD are in accordance with those from a case report evaluating EMD with neither etching nor chemical preparation of the root surfaces for the treatment of intrabony periodontal defects. At 12 months, the results demonstrated a mean PD reduction of  $4.4 \pm 1.3$  mm and a mean CAL gain of  $3.6 \pm 1.2$  mm.<sup>49</sup> These findings might also be supported by the results of a recent study, which failed to show statistically significant differences in terms of PD reduction and CAL gain following regenerative surgery with either EDTA+EMD or EMD alone.<sup>50</sup> All these data, taken together with the results of the present study seem to indicate that the clinical results may rather be attributed to the effect of EMD than to the root surface conditioning itself. In this context, however, it is important to realize that the presented clinical results need to be supported by his-

tologic evidence, since it is still unclear to what extent the CAL gains obtained following ERL+EMD represent real periodontal regeneration rather than defect fill without new connective tissue attachment. Furthermore, the stability of the obtained CAL gains over time must be evaluated in further clinical studies.

Within the limits of a case report study, it may be concluded that the combination of ERL and EMD may improve clinical healing of intrabony periodontal defects over a period of 6 months.

## REFERENCES

1. Claffey N, Polyzois I, Ziaka P. An overview of nonsurgical and surgical therapy. *Periodontol 2000* 2004;36:35-44.
2. Caton JG, Greenstein G. Factors related to periodontal regeneration. *Periodontol 2000* 1993;1:9-15.
3. Bowers GM, Chadroff B, Carnevale R, Mellonig J, Corio R, Emerson J, Stevens M, Romberg E. Histologic evaluation of new attachment apparatus formation in humans. Part I. *J Periodontol* 1989; 60:664-674.
4. Brunsvold MA, Mellonig JT. Bone grafts and periodontal regeneration. *Periodontol 2000* 1993;1:80-91.
5. Karring T, Lindhe J, Cortellini P. Regenerative periodontal therapy. *Clinical periodontology and implant dentistry* 3rd edition, pp 597-646 Copenhagen: Munksgaard, 1997.

6. Lowenguth RA, Blieden TM. Periodontal regeneration: root surface demineralization. *Periodontol* 2000 1993;1:54-68.
7. Lynch SE, Williams RC, Polson AM, Howell TH, Reddy MS, Zappa UE, Antoniadis HN. A combination of platelet-derived and insulin-like growth factors enhances periodontal regeneration. *J Clin Periodontol* 1989;16:545-548.
8. Hammarström L. Enamel matrix, cementum development and regeneration. *J Clin Periodontol* 1997;24:658-668.
9. Kawase T, Okuda K, Momose M, Kato Y, Yoshie H, Burns DM. Enamel matrix derivative (EMDOGAIN) rapidly stimulates phosphorylation of the MAP kinase family and nuclear accumulation of smad2 in both oral epithelial and fibroblastic human cells. *J Periodontol Res* 2001;36:367-376.
10. Kawase T, Okuda K, Yoshie H, Burns DM. Anti-TGF-beta antibody blocks enamel matrix derivative-induced upregulation of p21WAF1/cip1 and prevents its inhibition of human oral epithelial cell proliferation. *J Periodontol Res* 2002;37:255-262.
11. Suzuki S, Nagano T, Yamakoshi Y, Gomi K, Arai T, Fukae M, Katagiri T, Oida S. Enamel matrix derivative gel stimulates signal transduction of BMP and TGF-beta. *J Dent Res* 2005;84:510-514.
12. Takayama T, Suzuki N, Narukawa M, Tokunaga T, Otsuka K, Ito K. Enamel matrix derivative stimulates core binding factor alpha1/Runt-related transcription factor-2 expression via activation of Smad1 in C2C12 cells. *J Periodontol* 2005;76:244-249.
13. Hammarström L, Heijl L, Gestrelus S. Periodontal regeneration in a buccal dehiscence model in monkeys after application of enamel matrix proteins. *J Clin Periodontol* 1997;24:669-677.
14. Mellonig JT. Enamel matrix derivative for periodontal reconstructive surgery: technique and clinical and histologic case report. *Int J Periodontics Restorative Dent* 1999;19:8-19.
15. Sculean A, Donos N, Windisch P, Brex M, Gera I, Reich E, Karring T. Healing of human intrabony defects following treatment with enamel matrix proteins or guided tissue regeneration. *J Periodontol Res* 1999;34:310-322.
16. Yukna RA, Mellonig JT. Histologic evaluation of periodontal healing in humans following regenerative therapy with enamel matrix derivative. A 10-case series. *J Periodontol* 2000;71:752-759.
17. Sculean A, Windisch P, Chiantella GC, Donos N, Brex M, Reich E. Treatment of intrabony defects with enamel matrix proteins and guided tissue regeneration. A prospective controlled clinical study. *J Clin Periodontol* 2001;28:397-403.
18. Silvestri M, Ricci G, Rasperini G, Sartori S, Cattaneo V. Comparison of treatments of infrabony defects with enamel matrix derivative, guided tissue regeneration with a nonresorbable membrane and Widman modified flap. A pilot study. *J Clin Periodontol* 2000;27:603-610.
19. Heijl L, Heden G, Svardstrom G, Ostgren A. Enamel matrix derivative (EMDOGAIN) in the treatment of intrabony periodontal defects. *J Clin Periodontol* 1997;24:705-714.
20. Pontoriero R, Wennstrom J, Lindhe J. The use of barrier membranes and enamel matrix proteins in the treatment of angular bone defects. A prospective controlled clinical study. *J Clin Periodontol* 1999;26:833-840.
21. Blomlöf J, Lindskog S. Root surface texture and early cell and tissue colonization after different etching modalities. *Eur J Oral Sci* 1995;103:17-24.
22. Blomlöf JP, Blomlöf LB, Lindskog SF. Smear layer formed by different root planing modalities and its removal by an ethylenediaminetetraacetic acid gel preparation. *Int J Periodontics Restorative Dent* 1997;17:242-249.
23. Blomlöf JP, Blomlöf LB, Lindskog SF. Smear removal and collagen exposure after non-surgical root planing followed by etching with an EDTA gel preparation. *J Periodontol* 1996;67:841-845.
24. Polson AM, Frederick GT, Ladenheim S, Hanes PJ. The production of a root surface smear layer by instrumentation and its removal by citric acid. *J Periodontol* 1984;55:443-446.
25. Blomlöf J. Root cementum appearance in healthy monkeys and periodontitis-prone patients after different etching modalities. *J Clin Periodontol* 1996;23:12-18.
26. Blomlöf J, Blomlöf L, Lindskog S. Effect of different concentrations of EDTA on smear removal and collagen exposure in periodontitis-affected root surfaces. *J Clin Periodontol* 1997;24:534-537.
27. Gestrelus S, Andersson C, Johansson AC, Persson E, Brodin A, Rydhag L, Hammarström L. Formulation of enamel matrix derivative for surface coating. Kinetics and cell colonization. *J Clin Periodontol* 1997;24:678-684.
28. Aoki A, Sasaki KM, Watanabe H, Ishikawa I. Lasers in nonsurgical periodontal therapy. *Periodontol* 2000 2004;36:59-97.
29. Ishikawa I, Aoki A, Takasaki AA. Potential applications of Erbium:YAG laser in periodontics. *J Periodontol Res* 2004;39:275-285.
30. Aoki A, Ando Y, Watanabe H, Ishikawa I. In vitro studies on laser scaling of subgingival calculus with an erbium:YAG laser. *J Periodontol* 1994;65:1097-1106.
31. Eberhard J, Ehlers H, Falk W, Acil Y, Albers HK, Jepsen S. Efficacy of subgingival calculus removal with Er:YAG laser compared to mechanical debridement: an in situ study *J Clin Periodontol* 2003;30:511-518.
32. Schwarz F, Sculean A, Berakdar M, Szathmari L, Georg T, Becker J. In vivo and in vitro effects of an Er:YAG laser, a GaAlAs diode laser, and scaling and root planing on periodontally diseased root surfaces: a comparative histologic study. *Lasers Surg Med* 2003;32:359-366.
33. Buchanan SA, Robertson PB. Calculus removal by scaling/root planing with and without surgical access. *J Periodontol* 1987;58:159-163.
34. Hale GM, Querry MR. Optical constants of Water in the 200-nm to 200-µm wavelength region. *Appl Optics* 1973;12:555-563.
35. Schwarz F, Bieling K, Venghaus S, Sculean A, Jepsen S, Becker J. Influence of fluorescence-controlled Er:YAG laser radiation, the Vectortrade mark system and hand instruments on periodontally diseased root surfaces in vivo. *J Clin Periodontol* 2006;33:200-208.
36. Ando Y, Aoki A, Watanabe H, Ishikawa I. Bactericidal effect of erbium YAG laser on periodontopathic bacteria. *Lasers Surg Med* 1996;19:190-200.
37. Folwaczny M, Aggstaller H, Mehl A, Hickel R. Removal of bacterial endotoxin from root surface with Er:YAG laser. *Am J Dent* 2003;16:3-5.
38. Folwaczny M, Mehl A, Aggstaller H, Hickel R. Antimicrobial effects of 2.94 microm Er:YAG laser radiation on root surfaces: an in vitro study. *J Clin Periodontol* 2002;29:73-78.
39. Sugi D, Fukuda M, Minoura S, Yamada Y, Tako J, Miwa K, Noguchi T, Nakashima K, Sobue T, Noguchi T. Effects of irradiation of Er:YAG laser on quantity of endotoxin and microhardness of surface in exposed root after removal of calculus. *Jap J Cons Dent* 1998;41:1009-1017.
40. Yamaguchi H, Kobayashi K, Osada R, Sakuraba E, Nomura T, Arai T, Nakamura J. Effects of irradiation of an erbium:YAG laser on root surfaces. *J Periodontol* 1997;68:1151-1155.



41. Schwarz F, Sculean A, Georg T, Becker J. Clinical evaluation of the Er:YAG laser in combination with an enamel matrix protein derivative for the treatment of intrabony periodontal defects: a pilot study. *J Clin Periodontol* 2003;30:975-981.
42. Löe H. The Gingival Index, the Plaque Index and the Retention Index Systems. *Journal of Periodontology* 1967;38:Suppl:610-616.
43. Tonetti MS, Pini-Prato G, Cortellini P. Effect of cigarette smoking on periodontal healing following GTR in infrabony defects. A preliminary retrospective study. *J Clin Periodontol* 1995;22:229-234.
44. Schwarz F, Sculean A, Berakdar M, Georg T, Reich E, Becker J. Clinical evaluation of an Er:YAG laser combined with scaling and root planing for non-surgical periodontal treatment. A controlled, prospective clinical study. *J Clin Periodontol* 2003;30:26-34.
45. Schwarz F, Sculean A, Berakdar M, Georg T, Reich E, Becker J. Periodontal treatment with an Er:YAG laser or scaling and root planing. A 2-year-follow up-split mouth study. *J Periodontol* 2003; 74:590-596.
46. Schwarz F, Sculean A, Georg T, Reich E. Periodontal treatment with an Er: YAG laser compared to scaling and root planing. A controlled clinical study. *J Periodontol* 2001;72:361-367.
47. Folwaczny M, Thiele L, Mehl A, Hickel R. The effect of working tip angulation on root substance removal using Er:YAG laser radiation: an in vitro study. *J Clin Periodontol* 2001;28:220-226.
48. Froum SJ, Weinberg MA, Rosenberg E, Tarnow D. A comparative study utilizing open flap debridement with and without enamel matrix derivative in the treatment of periodontal intrabony defects: a 12-month re-entry study. *J Periodontol* 2001; 72: 25-34.
49. Parashis A, Tsiklakis K. Clinical and radiographic findings following application of enamel matrix derivative in the treatment of intrabony defects. A series of case reports. *J Clin Periodontol* 2000; 27: 705-713.
50. Sculean A, Berakdar M, Willershausen B, Arweiler N, Becker J, Schwarz F. Effect of EDTA Root Conditioning on the Healing of Intrabony Defects Treated with an Enamel Matrix Protein Derivative. *J Periodontol* 2006 (in press).

**Contact address:** Dr. Frank Schwarz, Department of Oral Surgery, Westdeutsche Kieferklinik, Heinrich Heine University, D-40225 Düsseldorf, Germany. Tel: +49-211-8118149, Fax: +49-211-1713542. e-mail: info@frank-schwarz.de